#### Lab 3: Short-read assembly: An overview

Maoying, Wu

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## What is assembly?



### Terminology and definitions

Fragment library a shrot insert (270bp) library with overlapping ends, a.k.a standard library

Long insert library A 4-8kb librbary where only 100 bps on each end are sequenced. a.k.a CLIP, mate pair library

Contig A contiguous sequence of DNA

Scaffold One or more contigs linked together by unknown sequence

Captured gap A gap within a scaffold. The order and orientation of the contigs spanning the gap is known.

# Traditional approach



- Sanger capillary sequencing often produce fragment of length 600 bp.
- $\sim 10x$  coverage / sequencing depth
- Assebled using overlap-layout-consensus approach.
  - Build an overlap graph where each node represents a read. An edge exists between two reads iff they overlap.
  - Traverse the graph to find unambiguous paths which form contigs.

## Next generation sequencing (NGS)

- Roche/454, Illumina Solexa, ABI SoLID
- Much higher throughput
- Lower cost
- Very short fragment lengths (25-300bp)
- Higher error rate
- Single-end or paired-end (mate-pair) sequencing

## Paired-end sequencing



- Sequencing two ends of a fragment of known size
- Currently fragment length (insert size) can range from 200 -10,000 bps

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## Assembly for NGS: Challenges

Challenges: impractical overlap-layout-consensus

- > 100x coverage, result in a large number of reads
- ► Short fragment → short overlaps will generate large fraction of false overlaps.

High sequencing error rate

#### Solutions:

- de Bruijin graph assembly
- Seed and extend assembly
- Velvet: the most popular short read assembly method.

## Current approach



## Genome assembly programs

Name	Algorithm	Data
Abyss	De Bruijn	Illumina
Allpaths-LG	De Bruijn	Illumina/PacBio
CABOG (Celera)	OLC	All
HGAP	OLC	PacBio
Masurca	De Bruijn/OLC	All
Mira	OLC	All
Newbler	OLC	454/Illumina/Torrent
SGA	String	Illumina
SoapDeNovo	De Bruijn	Illumina
Spades	De Bruijn	Illumina
Velvet	De Bruijn	Illumina

# De Bruijn graph method

#### Pros

high efficiency

#### Cons

- Partitioning all reads into k-mers is memory- intensive. For mammalian genomes, 1000+ GB RAM is prohibitively expensive.
- Loss of connectivity information between reads due to the chop of reads into k-mers. The longer the reads, the more information is lost.

## Assembly tips

- The structure of the graph is highly dependent on the k-mer size used for assembly.
  - Small k-mers result in shorter contigs but with lots of connections.
  - ► Large *k*-mers can result in longer contigs but with fewer connections.
- If your graph consists of many separate disconnected subgraphs, then your k-mer size may be too large.
- If your graph is connected but is very dense and tangled, then your k-mer size may be too small.
- When assembling 100 bp reads in Velvet, a k-mer of 51 would be a good starting point, and then adjust up or down as needed.
- SPAdes conveniently conducts assembly multiple times using different k-mers, so you can look at the FASTA files for each assembly (in folders named like K21, K33, etc.) to find the best graph for viewing in Bandage.

## Bandage: A GUI program to view assembly graphs

- De novo assembly graphs contain assembled contigs (nodes) but also the connections between those contigs (edges).
- Bandage visualizes assembly graphs, with connections, using graph layout algorithms.
- Users can interact with the graph by moving, labeling and coloring nodes.
- Sequence information can be directly extracted from the graph viewer.

## Install Bandage

- 1. sudo apt-get update
- sudo apt-get install build-essential git qtbase5-dev libqt5svg5-dev
- 3. Prepare the OGDF library:
  - Download OGDF code from http://www.ogdf.net and unzip
  - Create the Makefile: ./makeMakefile.sh
  - Compile the library: make
- Download the Bandage code: git clone https:// github.com/rrwick/Bandage.git
- 5. Set the environment variable: export QT\_SELECT=5
- 6. Run qmake to generate a Makefile: qmake
- 7. Build the program: make
- 8. You can install both OGDF and Bandage to /usr/local/bin
- 9. Run Bandage

### SGA: String graph assembler

► a.k.a. Simpson-Durbin assembler

- (1) computes overlaps between all read pairs
- (2) constructs a string graph based on the overlaps.
- (3) derives the genome assembly from the string graph.
- Pros:
  - considerably lower memory demand
  - handle huge genomes at a significantly lower cost.

Of the four assembler, SGA used the least memory (4.5 GB vs. 14.1 GB, 23.0 GB and 38.8 GB for ABySS, Velvet and SOAPdenovo, respectively)

### SGA: Reference

#### String Graph concept:

• E. W. Myers. The Fragment Assembly String Graph.

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